## Note

## Purification of inulobiose obtained by acid hydrolysis of inulin

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Inulobiose (1-O- $\beta$ -D-fructofuranosyl-D-fructose) was required in a pure state for mass spectrometry. Preparation of the disaccharide by partial acid hydrolysis of inulin (according to methods previously described<sup>1,2</sup>) yielded a product, isolated by paper chromatography, which gave 7.5% of D-glucose on acid hydrolysis. Inulobiose has an  $R_{\rm F}$  value very close to that of sucrose, and it was reasoned that the D-glucose must have arisen from contaminant sucrose.

Inulin has a d.p. of  $\sim$ 34 and the terminal residue is an  $\alpha$ -D-glucopyranose residue linked to a  $\beta$ -D-fructofuranose residue as in sucrose, the remainder of the molecule being composed of (2 $\rightarrow$ 1)-linked  $\beta$ -D-fructofuranose residues<sup>3</sup>. Since the rate constants for acid hydrolysis of inulobiose and sucrose, under the conditions described, are 0.018 and 0.042 min<sup>-1</sup>, respectively<sup>1</sup>, the ratio of sucrose to inulobiose in a partial hydrolysate of inulin should reduce to  $\sim$ 1:7, *i.e.*, a D-glucose content of 6.3%.

Paper electrophoresis of the impure inulobiose in a borate buffer removed the contaminant sucrose (migration rates towards the anode: sucrose -1.7, inulobiose +13.0 cm/h at 2 kV and 230-240 mamp). The ratio of the weights of the sucrose to inulobiose thus obtained was 1:6. The difference between this value and the theoretical ratio, together with the higher D-glucose content of the unresolved mixture (see above), suggests that the d.p. of the original inulin sample may have been <34. Samples of crude inulin, also tested, gave still higher ratios of sucrose to inulobiose.

Purified inulobiose had m.p. 189–192.5°,  $[\alpha]_D^{20}$  –66 ±0.5° (water), yielded no glucose on hydrolysis, and had a fructose content of >99%. The yield from 2 g of inulin was 25 mg.

The differences between the specific rotation of the purified product first recorded<sup>1</sup> and the values subsequently found<sup>2,4</sup> could partly be ascribed to the presence of sucrose in the first preparations. Moreover, the lower rotation of the samples prepared here suggests that some earlier preparations<sup>2,4</sup> may possibly have been contaminated with fructose.

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## EXPERIMENTAL

Inulin (2 g; Sigma Chemical Co.) was hydrolysed for 30 min at 70° in 10 ml of 0.01 M HCl; the resulting mixture was cooled and neutralised with sodium carbonate<sup>1</sup>. Alternatively, some hydrolysates were neutralised by passage through a column of Amberlite IR-4B(HO<sup>-</sup>) resin; on final purification, the inulobiose derived from these samples had a slightly higher (~1°) negative rotation. The neutralised solutions were concentrated to small volume under reduced pressure at 50°, and then subjected to chromatography<sup>5</sup> on Whatman 3MM paper using 7:1:2 and 6:1:3 1-propanol-ethyl acetate-water and 100:22:50 1-butanol-acetic acid-water in the descending mode. Fructosides were detected with urea-metaphosphoric acid<sup>6</sup>.

Electrophoresis was carried out in borate buffer<sup>7</sup>. The borate complexes derived from the electrophoresis were decomposed by evaporation of solutions at 40° in 9:1 methanol-water containing sufficient acetic acid to keep the pH at 3.5-4.0. Residual borate and other ions were removed by passing the product through Amberlite IR-120(H<sup>+</sup>) and IR-4B(HO<sup>-</sup>) resins. This last treatment was only carried out on samples that had previously been through the resin.

Total hydrolysis of the disaccharides was carried out in M H<sub>2</sub>SO<sub>4</sub> for 2 h at 100°; the hydrolysates were deionised by using resins as described above.

Fructose in solution was determined by the method of Cole (see ref. 8), and free D-glucose by the D-glucose oxidase method<sup>9</sup>.

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